

## Trunculins A and B, Norsesesterterpene Cyclic Peroxides from a Marine Sponge, *Latrunculia brevis*

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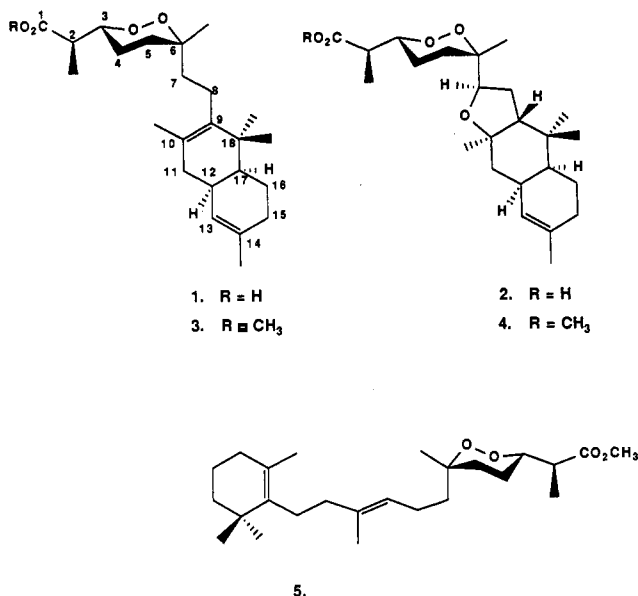
Received May 9, 1986

Two norsesesterterpene cyclic peroxides possessing a new carbon skeleton, trunculins A (1) and B (2), together with their methyl esters 3 and 4 have been isolated from a marine sponge, *Latrunculia brevis*. The esters 3 and 4 have been characterized and their relative stereostructures secured by detailed spectroscopic and X-ray analysis. Absolute stereochemistries are proposed on the basis of Horeau determinations carried out on the diol derivatives 11 and 12. The acids 1 and 2 exhibit antimicrobial activity.

In a recent report,<sup>1</sup> we described the isolation and structure elucidation of a number of antimicrobially active norterpene cyclic peroxides from two Australian sponges, a *Latrunculia* sps. and *Mycale (aegogrophila) cf. ancorina*. Another specimen from the same location, *Latrunculia brevis*,<sup>2</sup> has yielded two further examples of this class of marine natural product.

### Discussion

As with earlier studies, an ethanol extract of *L. brevis* was found to show significant growth-inhibitory activity against the Gram-positive bacteria *Bacillus subtilis* and the "yeast" *Saccharomyces cerevisiae*. Trituration of the concentrated extract with dichloromethane gave an antimicrobially active lipid-soluble material that on fractionation by rapid elution through silica yielded an active two-component mixture, trunculin A<sup>3</sup> (1) and trunculin B (2), together with an inactive mixture of the corresponding methyl esters 3 and 4. Although these could be resolved by reverse-phase HPLC, a more convenient separation was achieved by first methylating the crude extract with diazomethane followed by MPLC on silica, to yield<sup>4</sup> the monomethyl esters 3 and 4.



(1) Capon, R. J.; MacLeod, J. K. *Tetrahedron* 1985, 41, 3391.

(2) A specimen is lodged at the Northern Territory Museum of Arts and Sciences, Darwin, Australia, under the registry number Z4968.

(3) Assignment of the trivial name latrunculin to a series of unrelated marine natural products from *Latrunculia magnifica*<sup>13</sup> precludes its use here. Instead, the name trunculin has been coined to describe the natural products described herein.

(4) Both trunculin A (1) and its methyl ester 3 show characteristic bright green coloration on silica TLC when visualized with 5% vanillin in 3 M H<sub>2</sub>SO<sub>4</sub>. Trunculin B (2) and its methyl ester 4 exhibit an intense blue color under the same conditions.

Table I. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 50 MHz) Assignments for 3 and 4 and Selected Shifts for Model Compounds 5 and 10.

carbon	3	4	5 <sup>a</sup>	10 <sup>a,c</sup>
1	174.1 (s)	174.1 (s)	174.4 (s)	
2	42.6 (d)	42.6 (d)	42.6 (d)	
3	81.1 (d)	81.4 (d)	81.3 (d)	
4	22.5 (t)	22.4 (t)	22.6 (t)	
5	32.6 (t)	30.1 (t)	32.4 (t)	
6	79.9 (s)	80.7 (s)	79.9 (s)	
2-CH <sub>3</sub>	12.5 (q)	12.8 (q)	12.8 (q)	
6-CH <sub>3</sub>	23.6 (q)	20.6 (q)	23.9 (q)	
O-CH <sub>3</sub>	51.7 (q)	51.7 (q)	51.8 (q)	
7	34.9 (t)	73.1 (d)		
8	23.6 (t)	23.8 (t)		
9	137.6 (s)	50.9 (d)		
10	126.7 (s)	81.4 (s)		
11	40.1 (t)	41.7 (t)		
12	32.8 (d)	33.0 (d)		31.8 (d)
13	126.1 (d)	125.6 (d)		126.3 (d)
14	133.2 (s)	133.1 (s)		133.2 (s)
15	31.4 (t)	31.7 (t)		27.5 (t)
16	22.4 (t)	18.8 (t)		19.4 (t)
17	47.0 (d)	45.0 (d)		45.2 (d)
18	38.6 (s)	34.6 (s)		
10-CH <sub>3</sub>	19.9 (q)	28.6 <sup>b</sup> (q)		
14-CH <sub>3</sub>	23.3 (q)	23.1 (q)		23.5 (q)
18-CH <sub>3</sub>	21.6 (q)	19.0 (q)		
18-CH <sub>3</sub>	25.8 (q)	23.0 <sup>b</sup> (q)		

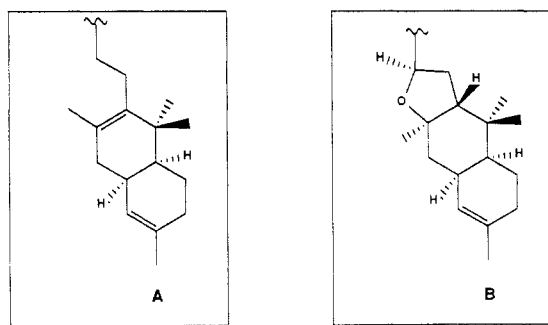
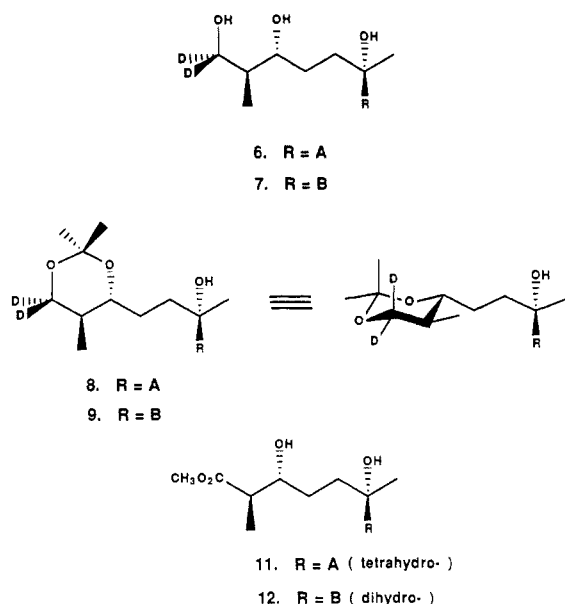
<sup>a</sup> Numbering assigned to model compounds 5 and 10 is as for 3 and 4. <sup>b</sup> May be interchanged. <sup>c</sup> Assignments made by current authors.

Table II. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 200 MHz) Assignments for 3 and 4

proton	3	4
2-H	2.56 (dq, 1 H, J = 8.0, 8.0 Hz)	2.51 (dq, 1 H, J = 8.0, 8.0 Hz)
3-H	4.24 (bddd, 1 H, J = 4.0, 8.0, 8.0 Hz)	4.25 (ddd, 1 H, J = 4.0, 8.0, 8.0 Hz)
2-CH <sub>3</sub>	1.14 (d, 3 H, J = 8.0 Hz)	1.14 (d, 3 H, J = 8.0 Hz)
6-CH <sub>3</sub>	1.16 (s, 3 H)	1.19 (s, 3 H)
O-CH <sub>3</sub>	3.69 (s, 3 H)	3.69 (s, 3 H)
7-H		4.54 (t, 1 H, J = 6.0, 6.0 Hz)
12-H		2.38 (br d, 1 H, J = 10.0 Hz)
13-H	5.20 (br s, 1 H)	5.34 (d, 1 H, J = 4.0 Hz)
15-H <sub>2</sub>		1.95 (br m, 2 H)
10-CH <sub>3</sub>	1.64 (s, 3 H)	0.90 (s, 3 H)
14-CH <sub>3</sub>	1.64 (s, 3 H)	1.63 (s, 3 H)
18-CH <sub>3</sub>	1.03 (s, 3 H)	0.90 (s, 3 H)
18-CH <sub>3</sub>	0.84 (s, 3 H)	0.99 (s, 3 H)

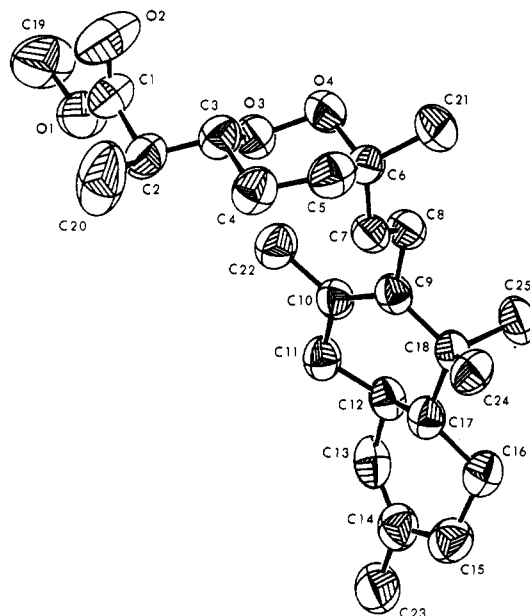
Comparison of the spectral data for 3 and 4 with that of known<sup>1</sup> norsesesterterpene cyclic peroxides, e.g. 5, confirmed the presence of the peroxide moiety (C1 to C6, Tables I and II). The equatorial nature of a carbon chain at C3 was apparent from the magnitude of the <sup>1</sup>H NMR

couplings to the C3 peroxy methine ( $J = 4, 8, \text{ and } 8 \text{ Hz}$ ) while the equatorial orientation of the C6-methyl could be ascertained from both  $^1\text{H}$  and  $^{13}\text{C}$  NMR shift data for **3**, although a definitive assignment could not be made for **4**.<sup>5</sup> It had previously been proposed<sup>1</sup> that the relative configuration between C2 and C3 in cyclic peroxy systems such as these could be assigned on the basis of the  $^1\text{H}$  NMR shift for the C2 secondary methyl (erythro  $\sim \delta 1.14$  and threo  $\sim \delta 1.25$ ). Thus it seemed likely that both trunculin A ( $\delta 1.14$ ) and trunculin B ( $\delta 1.14$ ) possessed an erythro configuration between C2 and C3. To confirm this, the cyclic peroxides **3** and **4** were converted via the triols **6** and **7** to the dioxans **8** and **9**. Both the  $^1\text{H}$  NMR shifts of the C2 secondary methyls ( $\delta 0.76$ , d,  $J = 7.0 \text{ Hz}$ ) and couplings to the C3 oxymethines (ddd,  $J = 2.0, 9.0, \text{ and } 9.0 \text{ Hz}$ ) in **8** and **9** were consistent<sup>1</sup> with an erythro configuration.

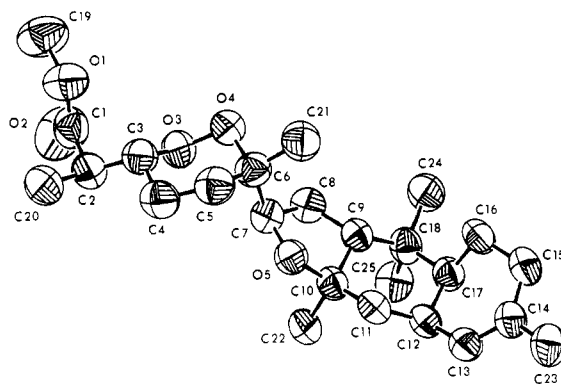


Also apparent from the  $^1\text{H}$  and  $^{13}\text{C}$  NMR analysis of **3** was the presence of a tri- and a tetrasubstituted double bond, two olefinic methyls, and a *gem*-dimethyl moiety. In the absence of other unsaturated functionalities, the unassigned portion of **3** was therefore bicyclic. Similarly, the unassigned portion of **4** possessed a trisubstituted double bond and a *gem*-dimethyl system. The presence of an additional oxygen atom and absence of a tetrasubstituted double bond in **4** relative to **3**, together with lack of an exchangeable proton or a ketone  $^{13}\text{C}$  NMR resonance, suggested the incorporation of a cyclic ether. Comparison

(5) In the case of **4**, the  $^{13}\text{C}$  NMR resonance attributed to the C6- $\text{CH}_3$  is at higher field than might otherwise be expected for an equatorial methyl in these systems, due to the influence of an adjacent five-membered ether moiety.

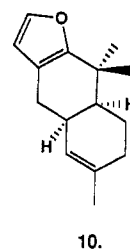


**Figure 1.** A view of molecule 1 of **3**,  $\text{C}_{25}\text{H}_{40}\text{O}_4$ . Thermal ellipsoids enclose 50% probability levels, and hydrogen atoms have been deleted for clarity, here and in Figure 2.



**Figure 2.** Thermal ellipsoid diagram of **4**,  $\text{C}_{25}\text{H}_{40}\text{O}_5$ .

of selected  $^{13}\text{C}$  NMR resonances in both **3** and **4** with those assigned to the carbocyclic system in furodysin<sup>6</sup> (**10**)



(Table I), together with one- and two-dimensional homo- and heteronuclear decoupling experiments, supported assignment of the gross structures as indicated. To confirm these assignments and to ascertain relative stereostructures for **3** and **4**, an X-ray analysis was undertaken.

Crystals of **3** and **4** suitable for X-ray analysis were obtained with difficulty from methanol/water. X-ray analysis of these crystals provided complete relative stereostructures for **3** and **4** (Figures 1 and 2),<sup>7</sup> which were fully consistent with the partial stereostructures proposed on the basis of spectroscopic evidence.

(6) Kazlauskas, R.; Murphy, P. T.; Wells, R. J.; Daly, J. J.; Schonholzer, P. *Tetrahedron Lett.* **1978**, 4951.

(7) Johnson, C. K. "ORTEP-II: A FORTRAN Thermal-Ellipsoid Plot Program for Crystal Structure Illustrations"; Report ORNL-5138; Oak Ridge National Laboratory: Oak Ridge, TN.

Absolute stereochemistries for trunculins A and B were determined by asymmetric esterification (Horeau analysis) of the saturated diol esters 11 and 12. In this manner the absolute stereochemistry about C3 was established as R, since both 11 and 12 returned a preponderance of (+)- $\alpha$ -phenylbutyric acid with optical yields of 9.3% and 4.8%, respectively. This procedure has previously been used<sup>1</sup> to assign absolute stereochemistries to marine norsterpene cyclic peroxides of this class.

### Experimental Section

**General Experimental.** <sup>1</sup>H NMR (200 MHz) and <sup>13</sup>C NMR (50 MHz) spectra were recorded on either a JEOL JNM-FX-200 or a Varian XL-200-E spectrometer. EIMS (70 eV) and CIMS (NH<sub>3</sub>) were obtained on a VG Micromass 7070F instrument. High-resolution accurate mass measurements were determined on an AEI MS 902 mass spectrometer. Optical rotations were recorded on a Perkin-Elmer 121 polarimeter. Antimicrobial activity was determined from a standard disc assay, with zones of inhibition being used as a measure of activity. For a description of the Horeau procedure, see ref 1.

*L. brevis* (Z4968) was collected by hand (SCUBA) at a depth of 20–25 m off South Durras, N.S.W., Australia. The fresh specimen was immersed in ethanol, packed in dry ice, transported to the laboratory, and stored at -20 °C. A portion of the EtOH extract was then examined by TLC, <sup>1</sup>H NMR spectroscopy, and antimicrobial assay.

The EtOH extract of Z4968 (50 g dry wt) after concentration to a brown gum (24 g) was triturated with CH<sub>2</sub>Cl<sub>2</sub> to give an antimicrobially active lipid-soluble fraction (3 g). Rapid silica filtration with stepwise gradient elution from hexane to EtOAc of a portion of this material (1 g) yielded an inactive fraction (20% EtOAc/hexane, 70 mg, 3 and 4) and an antimicrobially active fraction (EtOAc, 550 mg, 1 and 2). The latter material could be resolved by reverse-phase high-performance liquid chromatography (10- $\mu$ m C8 radial pak column eluted with 10% H<sub>2</sub>O/MeOH) into the crystalline solids trunculin A (1, mp 157–158 °C) and trunculin B (2, mp 96–98 °C). Methylation of 1 and 2 with diazomethane yielded monomethyl esters identical with those present (3 and 4) in the earlier inactive fractions. Thus a more convenient approach to purification involved methylation with diazomethane prior to rapid silica filtration, followed by elution through silica with 20% EtOAc/hexane to give a single inactive mixture of 3 and 4. This material was readily resolved by medium-pressure liquid chromatography (Lichroprep S<sub>1</sub> 60 (40–63  $\mu$ m) eluted with 20% Et<sub>2</sub>O/hexane) into the methyl esters of trunculins A (3) and B (4). Thus *L. brevis* (Z4968) contained four norsesterterpene cyclic peroxides, 1 (400 mg, 0.8%), 2 (1.22 g, 2.4%), 3 (100 mg, 0.2%), and 4 (110 mg, 0.22%), of which only the acids 1 and 2 exhibited antimicrobial activity.

**Trunculin A, methyl ester (3):** crystals from MeOH/H<sub>2</sub>O, mp 66.5–67.5 °C; [ $\alpha$ ]<sub>D</sub> +158.3° (c 1.02, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 200 MHz) (see Table II); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 50 MHz) (see Table I); EIMS, *m/z* 404.2915 (8%, M<sup>+</sup> requires C<sub>25</sub>H<sub>40</sub>O<sub>4</sub>, 404.2926), 389 (7), 386 (13), 373 (5), 335 (7), 317 (11), 299 (10), 203 (55), 202 (72), 201 (60), 187 (100), 81 (92).

**Trunculin B, methyl ester (4):** crystals from MeOH/H<sub>2</sub>O, mp 91–93 °C; [ $\alpha$ ]<sub>D</sub> +13.1° (c 1.54, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 200 MHz) (see Table II); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 50 MHz) (see Table I); EIMS, *m/z* 420 (~1%, M<sup>+</sup>), 405 (3), 402 (4), 389.2679 (5, M<sup>+</sup> - OCH<sub>3</sub> requires C<sub>24</sub>H<sub>37</sub>O<sub>4</sub>, 389.2692), 316.2399 (5, C<sub>21</sub>H<sub>33</sub>O<sub>2</sub> requires 316.2402), 233.1899 (100, C<sub>16</sub>H<sub>25</sub>O requires 233.1905).

**LiAlD<sub>4</sub> Reduction of 3.** To a solution of the cyclic peroxide 3 (26 mg, 0.06 mmol) in dry ether (3 mL) was added excess LiAlD<sub>4</sub> (10 mg, 0.24 mmol), and the resulting mixture was stirred under reflux for 2 h. The reaction was quenched by addition of 10% aqueous HCl (2 mL) and extracted with EtOAc. The EtOAc extract was then washed with H<sub>2</sub>O, dried with anhydrous MgSO<sub>4</sub>, and evaporated to yield the deuterated triol 6 (21 mg, 86%) as a stable colorless oil: <sup>1</sup>H NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$  0.83 (s, 3 H), 0.86 (d, 3 H, *J* = 7.0 Hz), 1.02 (s, 3 H), 1.24 (s, 3 H), 1.62 (s, 3 H), 1.66 (s, 3 H), 3.58 (m, 1 H), 5.19 (br s, 1 H); EIMS, *m/z* 362.3146 (1%, M<sup>+</sup> - H<sub>2</sub>O requires C<sub>24</sub>H<sub>38</sub>O<sub>2</sub>D<sub>2</sub>, 362.3154).

**LiAlD<sub>4</sub> Reduction of 4.** Reduction of 4 (17 mg) as described above for 3 yielded the deuterated triol 7 (14 mg, 87%) as a stable

colorless oil: <sup>1</sup>H NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$  0.85 (d, 3 H, *J* = 7.0 Hz), 0.90 (s, 3 H), 0.91 (s, 3 H), 1.16 (s, 3 H), 1.18 (s, 3 H), 1.63 (s, 3 H), 3.62 (br t, 1 H, *J* = 8.0 Hz), 3.85 (t, 1 H, *J* = 6.0 Hz), 5.34 (br d, 1 H, *J* = 4.0 Hz); EIMS, *m/z* 378.3110 (1%, M<sup>+</sup> - H<sub>2</sub>O requires C<sub>24</sub>H<sub>38</sub>O<sub>3</sub>D<sub>2</sub>, 378.3103), 363 (2), 317 (6), 145 (100).

**Isopropylidene Derivative 8.** The deuterated triol 6 (20 mg) in DMF (1 mL) was treated with 2,2-dimethoxypropane (0.2 mL) and *p*-toluenesulfonic acid (2 mg) and the resulting mixture stirred at room temperature overnight. The H<sub>2</sub>O-quenched reaction was then extracted with EtOAc to yield the isopropylidene derivative 8 (19 mg, 86%) as a stable colorless oil: <sup>1</sup>H NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$  0.76 (d, 3 H, *J* = 7.0 Hz), 0.84 (s, 3 H), 1.03 (s, 3 H), 1.20 (s, 3 H), 1.38 (s, 3 H), 1.43 (s, 3 H), 1.63 (s, 3 H), 1.66 (s, 3 H), 3.47 (ddd, 1 H, *J* = 2.0, 2.0, and 9.0 Hz), 5.19 (br s, 1 H); EIMS, *m/z* 405.3338 (13%, M<sup>+</sup> - CH<sub>3</sub> requires C<sub>26</sub>H<sub>41</sub>O<sub>3</sub>D<sub>2</sub>, 405.3338), 402 (16), 344 (12), 329 (10), 202 (80), 187 (65), 145 (71), 43 (100).

**Isopropylidene Derivative 9.** Treatment of the deuterated triol 7 (12 mg) as described above for 6 yielded the isopropylidene derivative 9 (11 mg, 83%) as a stable colorless oil: <sup>1</sup>H NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$  0.76 (d, 3 H, *J* = 7.0 Hz), 0.90 (s, 3 H), 0.91 (s, 3 H), 1.12 (s, 3 H), 1.18 (s, 3 H), 1.38 (s, 3 H), 1.42 (s, 3 H), 1.63 (s, 3 H), 3.46 (br t, 1 H, *J* = 9.0 Hz), 3.87 (t, 1 H, *J* = 6.0 Hz), 5.35 (br d, 1 H, *J* = 3.0 Hz); EIMS, *m/z* 421.3298 (5%, M<sup>+</sup> - CH<sub>3</sub> requires C<sub>26</sub>H<sub>41</sub>O<sub>4</sub>D<sub>2</sub>, 421.3287), 233 (11), 203 (27), 145 (100); CIMS (NH<sub>3</sub>), 437 (M + H, 50), 419, (55), 379 (80), 361 (100), 355 (66), 337 (61).

**Hydrogenation of 3.** A sample of the cyclic peroxide 3 (12 mg) in Et<sub>2</sub>O with 10% Pd/C catalyst (10 mg) was stirred under an atmosphere of H<sub>2</sub> for 4 h. The catalyst was removed by filtration through Celite and the product purified by stepwise elution (hexane to Et<sub>2</sub>O) through a silica sep-pak to give the saturated diol ester 11 (8 mg, 67%) as a stable colorless oil: <sup>1</sup>H NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$  0.68 (d, 3 H, *J* = 8.0 Hz), 0.72 (d, 3 H, *J* = 8.0 Hz), 0.88 (s, 3 H), 0.97 (d, 3 H, *J* = 8.0 Hz), 1.20 (s, 3 H), 1.23 (s, 3 H), 2.56 (dq, 1 H, *J* = 8.0 and 8.0 Hz), 3.71 (s, 3 H), 3.70 (br m, 1 H); EIMS, *m/z* 392.3291 (2%, M<sup>+</sup> - H<sub>2</sub>O requires C<sub>25</sub>H<sub>44</sub>O<sub>3</sub>, 392.3290), 377 (5), 374 (4), 359 (4), 305 (4), 261 (2), 191 (24), 171 (100).

**Hydrogenation of 4.** Treatment of 4 (19 mg) as described above for 3 yielded the saturated diol ester 12 (15 mg, 79%) as a stable colorless oil: <sup>1</sup>H NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta$  0.83 (s, 3 H), 0.85 (s, 3 H), 1.02 (d, 3 H, *J* = 6.0 Hz), 1.13 (s, 3 H), 1.16 (s, 3 H), 1.22 (d, 3 H, *J* = 6.0 Hz), 2.55 (dq, 1 H, *J* = 6.0 and 6.0 Hz), 3.71 (s, 3 H), 3.70 (br m, 1 H), 3.85 (br t, 1 H, *J* = 6.0 Hz); EIMS, *m/z* 406.3081 (1%, M<sup>+</sup> - H<sub>2</sub>O requires C<sub>25</sub>H<sub>42</sub>O<sub>4</sub>, 406.3083), 368 (4), 319 (3), 191 (5), 171 (100).

**Crystallography.** X-ray diffraction data were collected for compounds 3 and 4 on a Picker FACS-I diffractometer using graphite-monochromated Cu K $\alpha$  radiation. In each case, unit cell parameters were determined by least-squares analysis of the setting angles of 12 accurately centered reflections (80.6° < 2 $\theta$  < 85.7° for 3 and 86.4° < 2 $\theta$  < 96.1° for 4).  $\lambda$ (Cu K $\alpha$ ) = 1.5405 Å. Reflections with *I* > 3 $\sigma$ (*I*) were regarded as "observed" and were used in the structure solution and refinement. Data were corrected for anisotropic decay<sup>8</sup> and for absorption.<sup>9</sup> Both structures were solved by MULTAN<sup>10</sup> and the remaining non-hydrogen atoms located in difference electron-density maps. The chirality of the molecules was adjusted to give *S* configurations at C3 to be consistent with the Horeau determinations. Hydrogen atoms were positioned geometrically (*r*<sub>C-H</sub> = 0.95 Å, methyl groups derived from peaks in difference maps), *B*<sub>180</sub>(H) = 1.2 × *B*<sub>eq</sub>(adjacent C). Carbon and oxygen atoms were refined with anisotropic temperature factors, but hydrogen parameters were not varied. Block-diagonal least-squares analysis was used throughout for 3 in view of the large number of parameters, but full-matrix analysis was used in the final stages of refinement of 4. The function minimized in the least-squares analysis was  $\sum w(|F_o| -$

(8) Churchill, M. R.; Kalra, K. L. *Inorg. Chem.* 1974, 13, 1427.

(9) Sheldrick, G. M. SHELX-76, Program for Crystal Structure Determination; University of Cambridge, England.

(10) Main, P.; Fiske, S. J.; Hull, S. E.; Lessinger, L.; Germain, G.; Declercq, J. P.; Woolfson, M. M. MULTAN80. A System of Computer Programs for the Automatic Solution of Crystal Structures from X-ray Diffraction Data; Universities of York, England, and Louvain-la-Neuve, Belgium.

$|F_c|)^2$  where  $w = 1/[(\sigma(F))^2 + 0.25(0.05F)^2]$ . Neutral-atom scattering factors with anomalous dispersion corrections for oxygen and carbon were used throughout.<sup>11</sup> Computing<sup>12</sup> was carried out on a UNIVAC-1100/82 computer.

**Crystal Data. Compound 3:**  $2(C_{25}H_{40}O_4) \cdot 2.67H_2O$ ,  $M_r$  857.27, orthorhombic, space group  $P2_12_12_1$ ,  $a = 10.325$  (1) Å,  $b = 14.856$  (1) Å,  $c = 34.476$  (3) Å,  $V = 5288.2$  Å<sup>3</sup>,  $D_{\text{calcd}} = 1.079$  g cm<sup>-3</sup>,  $Z = 4$ ,  $\mu(\text{Cu K}\alpha) = 5.58$  cm<sup>-1</sup>. The crystal was colorless and of dimensions  $0.27 \times 0.32 \times 0.14$  mm.  $\theta$ - $2\theta$  scans were used to collect intensity data on 3753 unique reflections to  $2\theta_{\text{max}} 110^\circ$ , 3168 being "observed". The final  $R$  factor was  $R = 0.046$ ,  $R_w = 0.064$ . The maximum shift/error ratio was  $<0.4$  for "solvent molecules" and  $>0.2$  for other atoms. All features in a final difference map were  $<0.16$  e Å<sup>-3</sup>.

**Compound 4:**  $C_{25}H_{40}O_5$ ,  $M_r$  420.59, monoclinic, space group  $P2_1$ ,  $a = 11.553$  (1) Å,  $b = 9.759$  (1) Å,  $c = 11.900$  (2) Å,  $\beta = 114.56$  (1)°,  $V = 1220.3$  Å<sup>3</sup>,  $D_{\text{calcd}} = 1.144$  g cm<sup>-3</sup>,  $Z = 2$ ,  $\mu(\text{Cu K}\alpha) = 5.89$  cm<sup>-1</sup>. The crystal was colorless,  $0.18 \times 0.39 \times 0.17$  mm. Intensity data were collected by  $\omega$  scans to  $2\lambda_{\text{max}} 125^\circ$ , yielding 2091 unique reflections, 1856 "observed". Six reflections suffering from extinction were removed from the data set. The final  $R$  factor was

$R = 0.059$ ,  $R_w = 0.077$ . The maximum shift/error ratio was  $>0.1$ . All features in a final difference map were  $<0.19$  e Å<sup>-3</sup>.

Crystals of compound 3 were found to contain two independent molecules of  $C_{25}H_{40}O_4$ , showing essentially the same lengths and angles for equivalent bonds and very similar conformations, within the lattice. In addition, there were columns of electron density ( $\rho_{\text{max}} \sim 1.2$  e Å<sup>-3</sup>) running in the  $a$  direction through the crystal lattice, which would appear to correspond to disordered molecules of solvate, either  $H_2O$  or  $CH_3OH$ . Four oxygen atoms of occupancy 0.67 were introduced to model this region, though its exact nature was not determined.

**Acknowledgment.** Thanks go to Dr. E. Ball and Dr. R. Summons for assistance in specimen collection, to W. Wheate and M. Chapman for acquisition of mass spectral data, to J. Hooper for sponge taxonomy, and to M. Anderson and J. Rothschild for assistance in antimicrobial assaying of crude extracts and purified products.

**Registry No.** 1, 105969-64-0; 2, 105969-65-1; 3, 105969-66-2; 4, 105969-67-3; 6, 105969-68-4; 7, 105969-69-5; 8, 105969-70-8; 9, 105969-71-9; 11, 105969-72-0; 12, 105969-73-1; 2,2-dimethoxypropane, 77-76-9.

(11) *International Tables for X-ray Crystallography*; Kynoch: Birmingham, England, 1974; Vol. 4, pp 99, 149.

(12) McLaughlin, G. M.; Taylor, D.; Whimp, P. O. *The ANUCRYS Structure Determination Package*; Research School of Chemistry, Australian National University: Canberra, Australia.

(13) Kashman, Y.; Groweiss, A.; Shmueli, U. *Tetrahedron Lett.* 1980, 21, 3629.

**Supplementary Material Available:** Figure 3 (showing molecule 2 of 3) and Tables III-IX (atomic parameters and selected bond lengths and angles for 3 and 4) (20 pages). Ordering information is given on any current masthead page. Structure factor listings for 3 and 4 are available on request.

## $O^5$ -Methyl-(±)-(2'R,3'S)-psorospermin

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Received August 18, 1986

Psorospermin (1), a novel xanthone isolated from the ethanolic extract of the root of the woody African plant *Psorospermum febrifugum* Spach. (Guttiferae), has shown biological activity in the 9KB cell culture and in vivo P388 mouse leukemia systems, leading to further investigation of 1 as a potential antineoplastic agent. Following the reisolation, studies were initiated to determine the absolute stereochemistry and to complete the total synthesis of 1.  $O^5$ -Methyl-(±)-(2'R,3'S)-psorospermin (2) was chosen as our initial target to test the feasibility of the formation of the dihydrofuran and epoxide moieties in a stereoselective manner. The 4-allyl group of 7, which was formed by an ortho-Claisen rearrangement, was oxidized to aldehyde 9. A Wittig reaction gave the *E* allylic ester 10 as the predominant product. Reduction of 10 to 11, epoxidation, mesylation, and deblocking gave the free phenol 14. Compound 14 was cyclized with potassium *tert*-butoxide in one step to give (±)-2, an epimer of 1, demonstrating that the epoxydihydrofuran system could be constructed in a concerted and stereoselective manner and providing indirect proof of the absolute stereochemistry of 1.

Psorospermin (1) (NSC-266491) is an antitumor xanthone originally isolated by Kupchan and co-workers<sup>1</sup> as a result of an activity-directed fractionation of the ethanolic extract of the root of the African woody plant *Psorospermum febrifugum* Spach. (Guttiferae). The cytotoxicity of 1 in the in vitro 9KB cell culture system, and the significant activity in the in vivo P388 mouse leukemia system suggested further investigation of 1 as a potential antineoplastic agent. Research efforts in our laboratories have resulted in the reisolation<sup>2</sup> of 1 from *P. febrifugum*

and the recent assignment of the absolute stereochemistry<sup>3</sup> to be 2'R,3'R as shown in Figure 1. We report here the total synthesis of (±)-2, an epimer of 1, which serves both as a confirmation of the assignment of configuration of 1 and as a demonstration that the dihydrofuran and the epoxide moieties can be constructed in a concerted and stereoselective manner.<sup>4</sup>

Based on the retrosynthetic analysis in Figure 1, it is proposed that an intramolecular attack of an appropriate epoxide by a phenoxide ion would result in formation of

(1) Kupchan, S. M.; Strelman, D. R.; Sneden, A. T. *J. Nat. Prod.* 1980, 43, 296.

(2) Cassady, J. M.; Chang, C.-j.; Habib, A. M.; Ho, D.; Amonkar, A.; Masuda, S. In *Natural Products and Drug Development*, Alfred Benzon Symposium 20; Krosggaard-Larsen, P., Brogger Christensen, S., Kofod, H., Eds.; Munksgaard: Copenhagen, 1984; p 228.

(3) Habib, A. M.; Ho, D. K.; Masuda, S.; McCloud, T.; Reddy, K. S.; Aboushoer, M.; McKenzie, A.; Byrn, S. R.; Chang, C.-j.; Cassady, J. M., submitted for publication.

(4) Another synthesis resulting in  $O^5$ -methyl-(±)-1 and -(±)-2 was reported: Strelman, D. R. Ph.D. Thesis, University of Virginia, Charlottesville, Virginia, 1977.